



Additional data for the virtual bank of cytogenetics of Brazilian mammals: karyotypes of medium and large mammals

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Abstract: Cytogenetic studies are important for species characterization and differentiation; furthermore, due to the highly variable and complex characteristics, karyotypes can assist in research that seeks to detect changes in genome organization, reveal phylogenetic history, and distinguish cryptic species. From the articles published in the literature, the karyotype information of terrestrial mammals of medium and large sizes was collected, with the intention of gathering them in a virtual database. For the searches, tools such as PubMed (National Center for Biotechnology Information) and Google Scholar were used; the words “karyotype”, “diploid number” and “2n” were combined with the names of each species from the following orders of terrestrial mammals of Brazil: Cetartiodactyla, Carnivora, Cingulata, Lagomorpha, Perissodactyla, Pilosa and Primates. Here, we present information for 115 species with data referring to diploid number, the number of autosomes, the morphology of the sex chromosomes and, if present, karyotypic variation. From this information, Portable Document Format extension (.pdf) files were made for each species, which contained, as well as the aforementioned karyotype information, the name, author and year of publication of each species, diagrams from the original karyotypes and a link to download the original article. All documents and information are available at <http://citogenetica.ufes.br>, which already includes the cytogenetic data of Brazilian small mammals in its database.

Key-Words: Chromosome morphology; Database; Diploid number; Karyotype variation.

Resumo: Dados adicionais para o banco virtual de citogenética de mamíferos brasileiros: cariótipos de mamíferos de médio e grande portes. Estudos citogenéticos são importantes para a caracterização e diferenciação de espécies; além disso, devido às características altamente variáveis e complexas, os cariótipos podem auxiliar em pesquisas que buscam detectar mudanças na organização do genoma, revelar história filogenética e distinguir espécies crípticas. A partir dos artigos publicados na literatura, coletou-se a informação cariotípica de mamíferos terrestres de médio e grande porte, com o intuito de reuni-los em um banco de dados virtual. Para as buscas, foram utilizadas ferramentas como PubMed (Centro Nacional de Informações em Biotecnologia) e Google Scholar; palavras-chaves como “cariótipo”, “número diplóide” e “2n” foram combinadas com os nomes de cada espécie das seguintes ordens de mamíferos terrestres do Brasil: Cetartiodactyla, Carnivora, Cingulata, Lagomorpha, Perissodactyla, Pilosa e Primatas. Aqui, apresentamos informações para 115 espécies com dados referentes ao número diplóide, o número de braços autossômicos, a forma dos cromossomos sexuais e, se presente, a variação cariotípica. A partir dessas informações, foram confeccionados arquivos de extensão *Portable Document Format* (.pdf) para cada espécie, os quais continham, além do cariótipo, o nome, autor e ano de publicação de cada espécie, figuras vetorizadas dos cariótipos originais e um link para baixar o artigo original. Todos os documentos e informações estão disponíveis em <http://citogenetica.ufes.br>, que já inclui os dados citogenéticos de pequenos mamíferos brasileiros em sua base de dados.

Palavras-Chave: Banco de dados; Morfologia dos cromossomos; Número diplóide; Variação cariotípica.

INTRODUCTION

The Mammalia class is divided into 12 orders, of which seven are accounted for in Brazilian terrestrial mammals of medium and large sizes: Cetartiodactyla, with two families, six genera and 10 species; Carnivora, with seven families, 23 genera and 33 species; Cingulata,

with one family, five genera and 11 species; Lagomorpha, with one family, one genus and one species; Perissodactyla, with one family, one genus and one species; Pilosa, with four families, five genera and eight species; and Primates, with five families, 19 genera and 118 species (Paglia *et al.*, 2012). Most of these orders differ in relation to the number of species found in each of the



Brazilian biomes (Paglia *et al.*, 2012); they are species that play an important role in the dynamics of ecosystems, such as top predators of the food chain (Sazima *et al.*, 1982) and seed dispersers (Jordano *et al.*, 2006).

It is well known that cytogenetic data are important tools for taxonomic and systematic studies, therefore aiding in the diagnosis of morphologically similar species, since karyotypes may be species-specific (Azevedo *et al.*, 2012; Bonvicino & Weksler, 1998; Wurster & Benirschke 1968). Additionally, they contribute to the elucidation of evolutionary and phylogenetic ambiguities (Robinson & Yang, 2012), increasing knowledge of species relationships (Romanenko *et al.*, 2007; Voss & Jansa, 2009) and revealing changes in genome organization at intra- and interspecific levels (Marshall *et al.*, 2008; Sagrillo *et al.*, 2005).

Given the importance of cytogenetic studies and the high diversity of Brazilian mammals, the gathering of karyotype information into one virtual guide, to be frequently updated, will contribute as a consistent tool to assist researchers who could be interested in systematic studies of the evolution and taxonomy of this group. Thus, this study intended to gather the cytogenetic data available in the literature for terrestrial mammals of Brazil, in order to complement an existing virtual library (<http://citogenetica.ufes.br>), where karyotype data for about 180 species of small non-flying mammals in Brazil (Paresque *et al.*, 2018) are already available.

MATERIALS AND METHODS

To gather the karyotypes for each species, a survey of the scientific papers available in the literature was carried out. To do so, the search tools PubMed of the National Center for Biotechnology Information (NCBI) and Google Scholar were employed, using the keywords “karyotype”, “diploid number” and “2n”, combined with the names of the mammalian species of interest.

Once the cytogenetic data were obtained, a general spreadsheet was created in Excel® containing information such as: scientific and popular name of each species, 2n (diploid number), FN (fundamental number), sex chromosome morphology and references. From this information, files with a Portable Document File (.pdf) extension were made, which were then made available for consultation in the UFES website (<http://citogenetica.ufes.br>). On this site, the data were organized according to the current taxonomic classification for the group. Each of these files contain the name, author and year of publication of each species, their karyotypic data, vectorized images of the original karyotypes, references and a download link to the original paper (Figure 1).

RESULTS

The cytogenetic data of 115 species of terrestrial mammals in Brazil were collected, corresponding to the 63.2% of all species (Table 1). Among the orders that

present the known karyotypes of all the species are: Cetartiodactyla, Lagomorpha, Perissodactyla and Pilosa. Despite being the most diverse orders, Carnivora and Primates have scarce data, with only 39.4% and 61.8% of species, respectively, having known acknowledged karyotype information.

In Cetartiodactyla, the diploid number varied from $2n = 26$ and $FN = 46$ in *Tayassu pecari* to $2n = 70$ and $FN = 70$ in *Mazama gouazoubira*. *Mazama americana* was the species that presented the highest intraspecific karyotypic variation, ranging from $2n = 42$ to $2n = 53$, due to the occurrence of chromosomal rearrangements occurring in common ancestors (Almeida *et al.*, 2001). The high diversity of chromosome sets and variations in sex chromosomes, without a predominant morphology or size for X and Y, contributes to the identification of the species of this group, allowing for species-specific karyotypes.

For the order Carnivora, the karyotypes of 13 species were compiled. The diploid number ranged from $2n = 38$ in species of the families Felidae, Mustelidae and Procyonidae up to $2n = 76$ in *Atelocynus microtis* and *Chrysocyon brachyurus* (Family Canidae); the variation in autosome number ranged from $FN = 68$ for members of the families Mustelidae and Procyonidae to $FN = 106$ in *Cerdocyon thous*. It is worth mentioning that, in this case, the level of variation does not necessarily represent a high diversity of $2n$, because the diploid number within each family repeats, leading to low interspecific variation. Thus, the differentiation of the chromosomal complements is subject to the combination of the form and size of the autosomal and sex pairs of each species.

In Cingulata, information was recorded for nine species (81.8% of the total species present in Brazil), which showed a diploid number variation from $2n = 38$ in *Tolypeutes matacusa* to $2n = 64$ in *Dasybus* species. The autosomal number ranged from $FN = 68$ in *Dasybus septemcinctus* to $FN = 98$ in *Euphractus sexcinctus*. With regard to the morphology of sex chromosomes, the majority of the members of this group presented a submetacentric X and small acrocentric Y.

The Pilosa order presented a variation from $2n = 49$ in *Choloepus hoffmanni* to $2n = 65$ in *Choloepus didactylus*, whereas the variation in the autosomal number ranged from $FN = 56$ in *Bradypus tridactylus* to $FN = 108$ in members of the Myrmecophagidae family. As for the sex chromosomes, the meta/submetacentric form predominated for the X, with the acrocentric form of the Y in Myrmecophagidae and metacentric form in Bradypodidae.

Finally, for Primates, information was collected for 73 species, with distinct patterns of karyotype variations observed depending on the family. The Aotidae family presented a diploid number ranging from $2n = 46$ in *Aotus vociferans* to $2n = 54$ in *Aotus nancymae* and *A. trivirgatus*, with an X chromosome that showed a large submetacentric form for most species, and a Y that was variable. In Atelidae, variation ranged from $2n = 32$ in *Ateles paniscus* to $2n = 62$ in members of the genera *Brachyteles* and *Lagothrix*; the X chromosome was



CIMAB

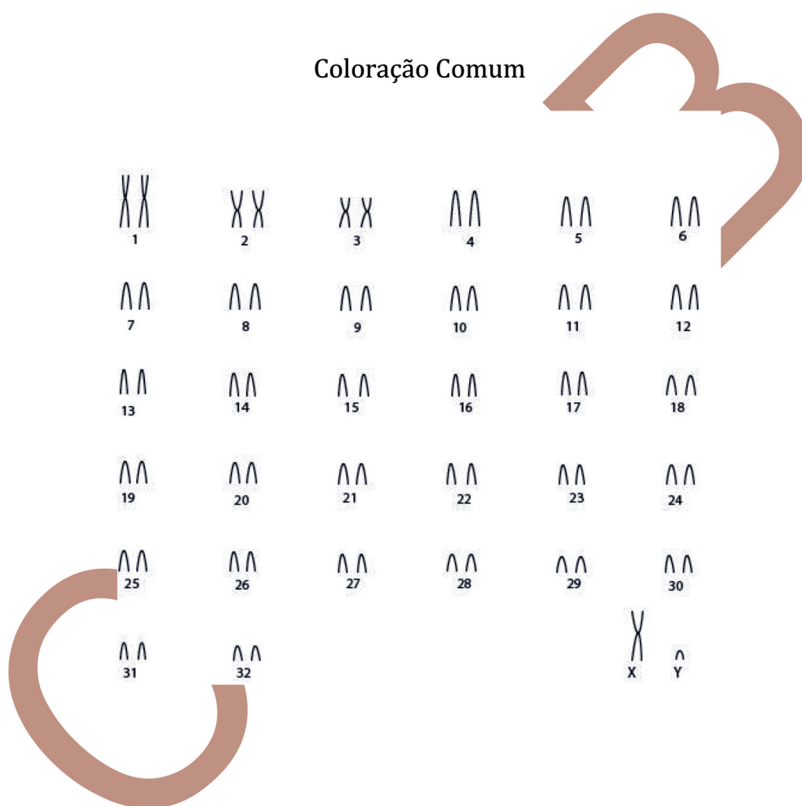
CITOGENÉTICA DE MAMÍFEROS DO BRASIL

Blastocerus dichotomus (Illiger, 1815)

Cariótipo de Duarte & Giannoni (1995)

2n	NF	Cromossomo X	Cromossomo Y	Localidade
66	74	Metacêntrico grande	Submetacêntrico pequeno	São Paulo (Brasil)

Coloração Comum



Referência

Barbanti Duarte, J. M.; Giannoni, M. L.. Cytogenetic analysis of the Marsh Deer, *Blastocerus dichotomus* (Mammalia, Cervidae). *Revista Brasileira de Genetica*, v. 18, n. 2, p. 245-248, 1995.

Link

<http://hdl.handle.net/11449/64568>

Figure 1: Model of the file prepared for each mammalian species of medium and large sizes whose karyotype is known in the literature. The files have a .pdf extension (Portable Document Format) and are available for consultation in the database at <http://citogenetica.ufes.br>. Each file presents: name, author and year of publication of each species, diploid number, autosomal number, form and size of the sex chromosomes, vectorized images of the original karyotypes, bibliographical reference and the hyperlink to consult the original article on the corresponding cytogenetic data.



Table 1: Compilation of cytogenetic data of medium and large Brazilian mammals, with the respective popular name in Brazil, diploid number (2n), fundamental number (FN), morphology of the X and Y chromosomes and references. ND = not described.

Order	Species	Popular name in Brazil	2n	FN	X	Y	References
Cetartiodactyla	<i>Blastocerus dichotomus</i>	Cervo do Pantanal	66	74	Large metacentric	Small submetacentric	Duarte & Giannoni, 1995
Cetartiodactyla	<i>Mazama americana</i>	Veado mateiro	42 to 53	48 to 56	Submetacentric	Metacentric	Cursino et al., 2014
Cetartiodactyla	<i>Mazama bororo</i>	Veado bororó de São Paulo	32 to 34	46	Medium submetacentric	Small acrocentric	Duarte & Jorge, 2003
Cetartiodactyla	<i>Mazama gouazoubira</i>	Veado catingueiro	70	70	Large Acrocentric	Small acrocentric	Neitzel, 1979
Cetartiodactyla	<i>Mazama nana</i>	Veado bororó do sul	36 to 40	56 to 60	Medium metacentric	Small metacentric	Abril & Duarte, 2008
Cetartiodactyla	<i>Mazama nemorivaga</i>	Veado da Amazônia	68-69	70	Medium submetacentric	Small metacentric	Fiorillo et al., 2013
Cetartiodactyla	<i>Odocoileus virginianus</i>	Veado da cara branca	70	74	Large submetacentric	Small metacentric	Wurster & Benirschke, 1967
Cetartiodactyla	<i>Ozotoceros bezoarticus</i>	Veado campeiro	68	74	Large metacentric	Small metacentric	Duarte & Giannoni, 1995
Cetartiodactyla	<i>Pecari tajacu</i>	Cateto or caítiu	30	46	Medium Acrocentric	Small acrocentric	Lima et al., 2004
Cetartiodactyla	<i>Tayassu pecari</i>	Queixada or porco do mato	26	46	Medium Acrocentric	Small acrocentric	Andrea et al., 2001; Adegá et al., 2007
Carnivora	<i>Atelocynus microtis</i>	Cachorro do mato de orelha curta	74 and 76	76	Submetacentric	Submetacentric	Wurster & Benirschke, 1968
Carnivora	<i>Cerdocyon thous</i>	Cachorro do mato	74	106	Large submetacentric	Small acrocentric	Hatanaka et al., 1998
Carnivora	<i>Chrysocyon brachyurus</i>	Lobo guará	76	78	Large submetacentric	Medium acrocentric	Pienkowska-Schelling et al., 2008
Carnivora	<i>Eira barbara</i>	Irara	38	68	Medium submetacentric	ND	Wurster & Benirschke, 1968
Carnivora	<i>Galictis cuja</i>	Furão	38	68	Metacentric	Small acrocentric	Barbosa, 2013
Carnivora	<i>Leopardus geoffroyi</i>	Gato do mato grande	38	72	Medium metacentric	Small submetacentric	Wurster & Benirschke, 1968
Carnivora	<i>Lycalopex gymnocercus</i>	Raposa do campo	74	76	Submetacentric	Small submetacentric	Wurster & Benirschke, 1968
Carnivora	<i>Lycalopex vetulus</i>	Raposinha	74	76	Metacentric	Acrocentric	Brum-Zorrilla & Langguth, 1980
Carnivora	<i>Nasua nasua</i>	Quati	38	68	Medium submetacentric	Metacentric	Wurster & Benirschke, 1968
Carnivora	<i>Panthera onca</i>	Onça pintada	38	72	Medium submetacentric	Small acrocentric	Wurster & Benirschke, 1968
Carnivora	<i>Pteronura brasiliensis</i>	Ariranha	38	64	Submetacentric	Submetacentric	Ledesma et al., 2004
Carnivora	<i>Puma yagouaroundi</i>	Gato mourisco	38	76	Medium submetacentric	ND	Franco-de-Sá et al., 2007
Carnivora	<i>Speothos venaticus</i>	Cachorro vinagre	74	ND	Large submetacentric	Small acrocentric	Wurster & Benirschke, 1968
Cingulata	<i>Cabassous tatouay</i>	Tatu de rabo mole grande	50	68	Small metacentric	Acrocentric	Barroso & Seuánez, 1991
Cingulata	<i>Cabassous unicinctus</i>	Tatu de rabo mole	46	ND	Medium submetacentric	Small acrocentric	Jacintho et al., 2009
Cingulata	<i>Dasyurus hybridus</i>	Tatu mulita	64	76 and 81	Large submetacentric	Small crocentric	Lizaralde et al., 2005; Saez et al., 1964
Cingulata	<i>Dasyurus kappleri</i>	Tatu de quinze quilos	64	ND	ND	ND	Redi et al., 2005
Cingulata	<i>Dasyurus novemcinctus</i>	Tatu galinha	64-65	78	Large metacentric	Small acrocentric	Beath et al., 1962; Bemirschke et al., 1969
Cingulata	<i>Dasyurus septemcinctus</i>	Tatui	64	76	Submetacentric	Acrocentric	Barroso & Seuánez, 1991
Cingulata	<i>Euphractus sexcinctus</i>	Tatu peba	58	98	Submetacentric or Acrocentric	Acrocentric	Jorge et al., 1978; Barroso & Seuánez, 1991
Cingulata	<i>Priodontes maximus</i>	Tatu canastra	50	76	Submetacentric	Metacentric	Bemirschke & Wurster, 1969
Cingulata	<i>Tolypeutes matacus</i>	Tatu bola	38	ND	Metacentric	Acrocentric	Jorge et al., 1978
Lagomorpha	<i>Sylvilagus brasiliensis</i>	Tapeti	40	68	Large submetacentric	Small submetacentric	Langguth & Sousa, 2008
Perissodactyla	<i>Tapirus terrestris</i>	Anta	80	80	Large submetacentric	Small acrocentric	Houck et al., 2000
Pilosa	<i>Bradypus torquatus</i>	Preguiça de coleira	50	64	Submetacentric	Metacentric	Azevedo et al., 2012
Pilosa	<i>Bradypus tridactylus</i>	Preguiça de três dedos	52	56	Metacentric	Metacentric	Dobigny et al., 2005
Pilosa	<i>Bradypus variegatus</i>	Bicho preguiça	54	ND	ND	ND	Azevedo et al., 2012
Pilosa	<i>Choloepus didactylus</i>	Preguiça real	65	ND	ND	ND	Dobigny et al., 2005



Order	Species	Popular name in Brazil	2n	FN	X	Y	References
Pilosa	<i>Choloepus hoffmanni</i>	Preguiça real	49	61	Metacentric	ND	Corin-Frederic, 1969
Pilosa	<i>Cyclopes didactylus</i>	Tamandú	64	100	Submetacentric	ND	Jorge et al., 1985; Jorge, 2000
Pilosa	<i>Myrmecophaga tridactyla</i>	Tamandú bandeira	60	108	Large metacentric	Small acrocentric	Pereira Júnior et al., 2004
Pilosa	<i>Tamandua tetradactyla</i>	Tamandú mirim	54	108	Large metacentric	Small acrocentric	Dobigny et al., 2005
Primate	<i>Alouatta belzebul</i>	Guariba-de-mãos-ruivas	49-50	70	Submetacentric	ND	Armada et al., 1987; Lima & Seuánez, 1989
Primate	<i>Alouatta caraya</i>	Bugio	52	70	Submetacentric	Acrocentric	Egozcue & De Egozcue, 1966
Primate	<i>Alouatta guariba</i>	Bugio ruivo	45-46, 49-50, 52	64, 66, 72	Submetacentric	Small acrocentric	Oliveira et al., 1998; Oliveira et al., 2000
Primate	<i>Alouatta macconnelli</i>	Bugio	47 to 49	ND	ND	ND	Lima & Seuánez, 1991
Primate	<i>Alouatta nigerima</i>	Bugio	50	66	Submetacentric	ND	Armada et al., 1987
Primate	<i>Alouatta seniculus</i>	Bugio	43 to 45	50, 52, 54	Small acrocentric	Small submetacentric	Yunis et al., 1976
Primate	<i>Alouatta ululata</i>	Guariba de mãos ruivas	49	70	Submetacentric	ND	Viana et al., 2015
Primate	<i>Aotus azarae</i>	Macaco da noite	49-50	69-70	Metacentric	ND	Mudry de Pargament et al., 1984
Primate	<i>Aotus fulvatus</i>	Macaco da noite	49-50	ND	ND	ND	Pieczarka & Nagamachi, 1988
Primate	<i>Aotus nancymae</i>	Macaco da noite	54	72	Large submetacentric	Small acrocentric	Pieczarka et al., 1992
Primate	<i>Aotus nigriceps</i>	Macaco da noite	51-52	65-66	Submetacentric	Submetacentric	Ma et al., 1980
Primate	<i>Aotus trivirgatus</i>	Macaco da noite	50 and 54	72 and 74	Submetacentric	Small metacentric	Egozcue et al., 1969; Menezes et al., 2010
Primate	<i>Aotus vociferans</i>	Macaco da noite	46 to 48	70	Large submetacentric	Small acrocentric	Pieczarka et al., 1992; Ma et al., 1976; Descailleux et al., 1990
Primate	<i>Ateles belzebul</i>	Macaco aranha	34	62	Medium submetacentric	Acrocentric	Medeiros et al., 1997
Primate	<i>Ateles chamek</i>	Macaco aranha da cara preta	34	62	Medium submetacentric	Small acrocentric	Medeiros et al., 1997
Primate	<i>Ateles marginatus</i>	Macaco aranha	34	62	Medium submetacentric	Subtelocentric	Medeiros et al., 1997
Primate	<i>Ateles paniscus</i>	Macaco aranha	32	58	Metacentric	Acrocentric	Pieczarka et al., 1989
Primate	<i>Brachyteles arachnoides</i>	Muriqui do sul	62	78	Submetacentric	ND	Koiffman & Saldanha, 1978
Primate	<i>Cacajao calvus</i>	Uacari-branco	46	ND	Submetacentric	Acrocentric	Bernirschke et al., 1976
Primate	<i>Cacajao melanocephalus</i>	Uacari-branco	45	65	Submetacentric	Acrocentric	Koiffmann & Saldanha, 1981
Primate	<i>Callicebus coimbrai</i>	Guigó	44	ND	ND	ND	Rodrigues et al., 2004
Primate	<i>Callicebus cupreus</i>	Zogue-zogue	46	66	Submetacentric	Small acrocentric	Dumas et al., 2005
Primate	<i>Callicebus donacophilus</i>	Zogue-zogue	50	70	Submetacentric	Submetacentric	Minezawa & Borda, 1984; de Boer, 1974; Barros et al., 2003
Primate	<i>Callicebus hoffmannsi</i>	Zogue-zogue	50	68	Submetacentric	Acrocentric	Rodrigues et al., 2001
Primate	<i>Callicebus lugens</i>	Zogue-zogue	16	22	ND	ND	Bonvicino et al., 2000; Stanyon et al., 2003
Primate	<i>Callicebus moloch</i>	Arabasu	46, 48 e 50	64, 66 e 72	Submetacentric	Small acrocentric	Dumas et al., 2005
Primate	<i>Callicebus pallescens</i>	Guigó	50	66	Submetacentric	Metacentric	Dumas et al., 2005
Primate	<i>Callicebus personatus</i>	Guigó	44	62	Submetacentric	ND	Rodrigues et al., 2004
Primate	<i>Callicebus personatus nigrifrons</i>	Guigó	42	68	Submetacentric	Metacentric	Nagamachi et al., 2003
Primate	<i>Callicebus torquatus</i>	Zogue-zogue	20 and 22	26 and 28	Submetacentric	ND	Egozcue et al., 1969; Bernirschke & Bogart, 1976; Barros et al., 2000
Primate	<i>Callimico goeldii</i>	Sagui de Goeldi	47-48	ND	Submetacentric	Small acrocentric	Dutrillaux et al., 1988
Primate	<i>Callithrix aurita</i>	Sagui da serra escuro	46	74	Submetacentric	Acrocentric	Nagamachi et al., 1997
Primate	<i>Callithrix flaviceps</i>	Sagui da serra	46	ND	ND	ND	Armada et al., 1982
Primate	<i>Callithrix geoffroyi</i>	Sagui de cara branca	46	74	Medium submetacentric	Metacentric	Nagamachi et al., 1997



Order	Species	Popular name in Brazil	2n	FN	X	Y	References
Primate	<i>Callithrix jacchus</i>	Sagui de tufo branco	46	74	Medium submetacentric	Acrocentric	Mudry et al., 1990; Sherlock, 1996
Primate	<i>Callithrix kuhlii</i>	Sagui	46	74	Medium submetacentric	Metacentric	Nagamachi et al., 1997
Primate	<i>Callithrix penicillata</i>	Sagui	46	74	Medium submetacentric	Metacentric or submetacentric	Nagamachi et al., 1997
Primate	<i>Cebuella pygmaea</i>	Sagui-leãozinho	44	74	Submetacentric	Small acrocentric	Nagamachi et al., 1992
Primate	<i>Cebus albifrons</i>	Caiarara	52 and 54	68	Submetacentric	Small acrocentric	Amaral et al., 2008; Egozcue e Egozcue, 1967
Primate	<i>Cebus olivaceus</i>	Caiarara	52	70	Medium submetacentric	Small acrocentric	Amaral et al., 2008
Primate	<i>Chiropotes israelita</i>	Cuxiú	54	74	Submetacentric	ND	Stanoyne et al., 2004
Primate	<i>Chiropotes satanas chiropotes</i>	Cuxiú	54	74	Submetacentric	Acrocentric	Seuáñez et al., 1992
Primate	<i>Chiropotes satanas utahicki</i>	Cuxiú	54	74	Submetacentric	Acrocentric	Seuáñez et al., 1992
Primate	<i>Lagothrix lagothricha</i>	Macaco barrigudo	62	88	Medium submetacentric	Small acrocentric	Stanyon et al., 2001; Clemente et al., 1987
Primate	<i>Lagothrix lagothricha cana</i>	Macaco barrigudo	62	90	Medium submetacentric	Small acrocentric	García et al., 1980
Primate	<i>Lagothrix lagothricha poeppigii</i>	Macaco barrigudo prateado	62	90	ND	ND	García et al., 1980
Primate	<i>Leontopithecus caissara</i>	Mico-leão-de-cara-preta	46	74	Submetacentric	Acrocentric	Sbalqueiro et al., 1992
Primate	<i>Leontopithecus chrysomelas</i>	Mico-leão-de-cara-dourada	46	74	Submetacentric	Subtelocentric	Nagamachi et al., 1997
Primate	<i>Leontopithecus chrysopygus</i>	Mico-leão-preto	46	76	Submetacentric	Subtelocentric	Seuáñez et al., 1988
Primate	<i>Leontopithecus rosalia</i>	Mico-leão-dourado	46	74	Submetacentric	Subtelocentric	Nagamachi et al., 1997
Primate	<i>Mico argentatus</i>	Sagui-branco	44	74	Submetacentric	Small metacentric	Egozcue et al., 1968
Primate	<i>Mico chrysolaucus</i>	Sauim	44	74	Submetacentric	Small metacentric	Nagamachi et al., 1996
Primate	<i>Mico emiliae</i>	Sauim	44	74	Submetacentric	Small metacentric	Barros et al., 1990
Primate	<i>Mico humeralifer</i>	Sagui	44	74	Submetacentric	Small metacentric	Nagamachi et al., 1996
Primate	<i>Mico mauesi</i>	Sagui-de-Maués	44	74	Large submetacentric	Small acrocentric	Nagamachi et al., 1994
Primate	<i>Pithecia irrorata</i>	Parauacú	48	64	Medium submetacentric	ND	Finotelo et al., 2010
Primate	<i>Pithecia pithecia</i>	Parauacú	48	64	Metacentric	Small acrocentric	Henderson et al., 1977
Primate	<i>Saguinus bicolor</i>	Sauim-de-coleira	46	74	Submetacentric	Metacentric or acrocentric	Dantas & Barros, 1997
Primate	<i>Saguinus fuscicollis</i>	Sauim-de-cara-suja	46	74	Submetacentric	Small metacentric	Egozcue et al., 1969
Primate	<i>Saguinus imperator</i>	Sauim-imperador	46	74	Submetacentric	Acrocentric	Dantas & Barros, 1997
Primate	<i>Saguinus labiatus</i>	Sauim-de-bigode	46	74	Submetacentric	Acrocentric	Marczynska et al., 1983
Primate	<i>Saguinus martinsi</i>	Sauim	46	74	ND	ND	Dantas & Barros, 1997
Primate	<i>Saguinus midas</i>	Sagui-de-mão-dourada	46	74	Submetacentric	Acrocentric	Nagamachi & Pieczarka, 1988
Primate	<i>Saguinus mystax</i>	Sagui-de-boca-branca	46	74	Submetacentric	Metacentric	Dantas & Barros, 1997
Primate	<i>Saguinus niger</i>	Sagui-una	46	74	Submetacentric	ND	Nagamachi & Pieczarka, 1988
Primate	<i>Saguinus nigricollis</i>	Sagui, sauim	46	74	Submetacentric	Metacentric	Benirschke et al., 1962
Primate	<i>Saguinus oedelli</i>	Sauim	46	74	Submetacentric	Acrocentric	Dantas & Barros, 1997
Primate	<i>Saimiri boliviensis</i>	Macaco-de-cheiro	44	72	Medium submetacentric	Small acrocentric	García et al., 1995
Primate	<i>Saimiri macdonon</i>	Macaco-de-cheiro	44	74	Submetacentric	Acrocentric	Bender & Mettler, 1958
Primate	<i>Saimiri sciureus</i>	Macaco-de-cheiro	44	74	Submetacentric	Acrocentric	Mudry et al., 1990
Primate	<i>Saimiri ustus</i>	Macaco-de-cheiro	44	74	ND	ND	Romagnolo, 2001
Primate	<i>Saimiri vanzolinii</i>	Macaco-de-cheiro	44	72	ND	ND	Yassuda & Chu, 1985
Primate	<i>Sapajus apella</i>	Macaco-prego	54	72	Submetacentric	Submetacentric	Freitas & Seuáñez, 1981



predominantly submetacentric, but variable in size, and the Y was as variable in form as in size. The Callitrichidae family presented low interspecific variation, from $2n = 44$ in species of *Cebuella* and *Mico* to $2n = 48$ in *Callimico goeldii*; for this group, an $FN = 74$ predominated, with the exception of *Leontopithecus chrysopygus*, which presented an $FN = 76$. The same constancy was observed in the submetacentric form of the X chromosome, with having only the Y chromosome showed any variation. In Cebidae, the diploid number ranges from $2n = 44$ in members of the genus *Saimiri* to $2n = 54$ in *Sapajus paella*; the submetacentric forms for the X and acrocentric for the Y prevailed in the sex chromosomes. Representatives of Pitheciidae presented the greatest variation, ranging from $2n = 16$ and $FN = 22$ in *Callicebus lugens* to $2n = 54$ and $FN = 74$ in members of the genus *Chiropotes*; the X chromosome was shown to be submetacentric, except for in *Pithecia pithecia*, while different forms and sizes of the Y chromosome were observed. In general, there is a consensus among the authors about the importance of cytogenetic data as a tool that is capable of assisting in the diagnosis of primate species.

From this information, .pdf files were made for all species, which contained images of the karyotypes recorded in the literature (Figure 1); these files are available and can be consulted at <http://citogenetica.ufes.br>.

DISCUSSION

The database of Cytogenetics of Brazilian Mammals (CIMAB) and its electronic address are available for consultation. This compilation is the result of a constant effort and is an important step in facilitating access to the cytogenetic characteristics of mammalian species occurring in Brazil. Some taxonomic groups, such as Carnivora and Primates, still lack karyotype data for a number of species; these orders were the ones with the lowest number of species with known cytogenetic data, indicating that new efforts should be made, focusing on these groups. The combined analysis of diploid numbers, autosomal numbers, as well as size and shape of the sex chromosomes at different taxonomic levels revealed that cytogenetics can be considered an important tool for the recognition and diagnosis of species, since, for the most part, this combination leads to karyotypes that are considered species-specific. The work of implanting the site and the insertion of new data should not stop here; the team from the Research Center on Evolution and Anatomy is constantly gathering efforts to keep the information updated with the publication of new articles. In addition, all researchers and authors of articles on mammalian cytogenetics are invited to contribute with new information. In the future, we intend to add information to other groups of mammals that have not yet been contemplated, along with data on interdisciplinary techniques involving chromosomes; our intention is to build a solid database and reference for the study of mammalian cytogenetics.

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REFERENCES

- Abril VV, Duarte JMB. 2008. Chromosome polymorphism in the Brazilian dwarf brocket deer, *Mazama nana* (Mammalia, Cervidae). *Genetics and Molecular Biology* 31(1): 53-57. <http://doi.org/10.1590/S1415-47572008000100011>.
- Adega F, Chaves R, Guedes-Pinto H. 2007. Chromosomal evolution and phylogenetic analyses in *Tayassu pecari* and *Pecari tajacu* (Tayassuidae): tales from constitutive heterochromatin. *Journal of Genetics* 86(1): 19-26. <http://doi.org/10.1007/s12041-007-0003-1>.
- Almeida CAS, Bonvicino C, Lachtermacher M, Moreira MAM, Olício R, Seuánez HN. 2001. Técnicas de avaliação da diversidade genética. Pp. 268-294. In: Garay I, Dias B (Eds.), *Conservação da biodiversidade em ecossistemas tropicais*. Vozes, Petrópolis.
- Amaral PJS, Finotelo LFM, Oliveira EHC, Pissinatti A, Nagamachi CY, Pieczarka JC. 2008. Phylogenetic studies of the genus *Cebus* (Cebidae: Primates) using chromosome painting and G-banding. *BMC Evolutionary Biology* 8(1): 169. <http://doi.org/10.1186/1471-2148-8-169>.
- Andrea MV, Oliveira C, Rocha GT, Foresti F. 2001. Cytogenetical and histological studies in testis of *Tayassu tajacu* (Cateto), *Tayassu pecari* (Queixada) and a natural interspecific hybrid. *Journal of Animal Breeding and Genetics* 118(2): 125-134. <http://doi.org/10.1046/j.1439-0388.2001.00275.x>.
- Armada JL, Barroso CM, Lima M, Muniz JAP, Seuánez HN. 1987. Chromosome studies in *Alouatta belzebul*. *American Journal of Primatology* 13(3): 283-296. <http://doi.org/10.1002/ajp.1350130306>.
- Armada JL, Soares VM, Seuánez H. 1982. Chromosome number in *Callithrix jacchus flaviceps* (Thomas, 1903). *Laboratory Primate Newsletter* 21: 1-3.
- Assis MFL, Barros R. 1987. Karyotype pattern of *Saimiri ustus*. *International Journal of Primatology* 8(5): 552-552.
- Azevedo NF, Svartman M, Manchester A, de Moraes-Barros N, Stanyon R, Vianna-Morgante AM. 2012. Chromosome painting in three-toed sloths: a cytogenetic signature and ancestral karyotype for Xenarthra. *BMC Evolutionary Biology* 12(1): 36. <http://doi.org/10.1186/1471-2148-12-36>.
- Barbosa LT. 2013. Contribuições citogenéticas nas espécies *Galictis cuja* (Carnivora, Mustelidae) e *Scapteromys* (Rodentia, Cricetidae): 1. Descrição cromossômica, utilizando citogenética clássica e molecular, da espécie *Galictis cuja* (Carnivora, Mustelidae) 2. Caracterização cariotípica através da citogenética clássica e análise filogenética em citótipos do gênero *Scapteromys* (Rodentia, Cricetidae). Dissertação em Genética, Programa de Pós-Graduação em Genética, Universidade Federal do Paraná, Curitiba, Brasil.
- Barros RMS, Nagamachi CY, Pieczarka JC. 1990. Chromosomal evolution in *Callithrix emiliae*. *Chromosoma* 99(6): 440-447. <http://doi.org/10.1007/BF01726697>.
- Barros RMS, Nagamachi CY, Pieczarka JC, Rodrigues LRR, Neusser M, Oliveira EH, Wienberg J, Muniz JAPC, Rissino JD, Muller S. 2003. Chromosomal studies in *Callicebus donacophilus pallascens*, with classic and molecular cytogenetic approaches: multicolour FISH using human and *Saguinus oedipus* painting probes. *Chromosome Research* 11(4): 327-334. <http://doi.org/10.1023/A:1024039907101>.
- Barros R, Pieczarka JC, Brigido MDCO, Muniz JAP, Rodrigues LR, Nagamachi CY. 2000. A new karyotype in *Callicebus torquatus* (Cebidae: Primates). *Hereditas* 133(1): 55-58. <http://doi.org/10.1111/j.1601-5223.2000.t01-1-00055.x>.
- Barroso CM, Seuánez H. 1991. Chromosome studies on *Dasyypus*, *Euphractus* and *Cabassous* genera (Edentata: Dasyproctidae). *Cytobios* 68(274-275): 179-196.



- Beath MM, Benirschke K, Brownhill LE. 1962. The chromosomes of the nine-banded armadillo, *Dasypus novemcinctus*. *Chromosoma* 13(1): 27-38. <http://doi.org/10.1007/BF00349617>.
- Bender MA, Mettler LE. 1958. Chromosome studies of primates. *Science* 128(3317): 186-190. <http://www.jstor.org/stable/1754678>.
- Benirschke K, Boer LEM, Bogart M. 1976. The karyotypes of two uakari species, *Cacajao calvus* and *C. rubicundus*. *Primates, Platyrrhini. Genen Phaenen* 19: 1-6.
- Benirschke K, Bogart MH. 1976. Chromosomes of the tan-handed titi (*Callicebus torquatus*, Hoffmannsegg, 1807). *Folia Primatologica* 25(1): 25-34. <http://doi.org/10.1159/000155705>.
- Benirschke K, Brownhill LE. 1962. Further observations on marrow chimerism in marmosets. *Cytogenetic and Genome Research* 1(5): 245-257. <http://doi.org/10.1159/000129734>.
- Benirschke K, Low RJ, Ferm VH. 1969. Cytogenetic studies of some armadillos. *Comparative Mammalian Cytogenetics* 330-345. http://doi.org/10.1007/978-3-642-85943-4_20.
- Bonvicino CR, Penna-Firme V, do Nascimento FF, Lemos B, Stanyon R, Seuánez HN. 2003. The lowest diploid number ($2n = 16$) yet found in any primate: *Callicebus lugens* (Humboldt, 1811). *Folia Primatologica* 74(3): 141-149. <http://doi.org/10.1159/000070647>.
- Bonvicino CR, Weksler M. 1998. A new species of *Oligoryzomys* (Rodentia, Sigmodontinae) from northeastern and central Brazil. *Zeitschrift für Säugetierkunde* 63(2): 90-103.
- Brum-Zorrilla N, Langguth A. 1980. Karyotype of South American pampas of *Pseudalopex gymnocercus* (Carnivora, Canidae). *Experientia* 36(9): 1043-1044. <http://doi.org/10.1007/BF01965957>.
- Carvalho TDL, Curi RA, Santiloni V, Chierregatto CAFDS, Rocha GT, da Mota LSL. 2010. Cytogenetic and molecular characterization of *Speothos venaticus* specimens. *Acta Scientiarum: Biological Sciences* 397-402. <http://hdl.handle.net/11449/42714>.
- Christoff JD, Kelly JS, Georgakis EG. 2000. U.S. Patent No. 6,071,119. Washington, DC: U.S. Patent and Trademark Office.
- Clemente IC, Garcia M, Ponsá M, Egozcue J. 1987. High-resolution chromosome banding studies in *Cebus apella*, *Cebus albifrons*, and *Lagothrix lagothricha*: Comparison with the human karyotype. *American Journal of Primatology* 13(1): 23-36. <http://doi.org/10.1002/ajp.1350130105>.
- Corin-Frederic J. 1969. So-called aberrant gonosomal formulas in placental mammals. The special case of the sloth *Choloepus hoffmanni* Peters (Edentata, Xenartha, Family Bradypodidae). *Chromosoma* 27(3): 268.
- Cursino MS, Salviano MB, Abril VV, dos Santos Zanetti E, Duarte JMB. 2014. The role of chromosome variation in the speciation of the red brocket deer complex: the study of reproductive isolation in females. *BMC Evolutionary Biology* 14(1): 40. <http://doi.org/10.1186/1471-2148-14-40>.
- Dantas SMMDM, Barros RMDS. 1997. Cytogenetic study of the genus *Saguinus* (Callithricidae, Primates). *Brazilian Journal of Genetics* 20(4). <http://doi.org/10.1590/S0100-84551997000400014>.
- Descailleaux J, Fujita R, Rodriguez LA, Aquino R, Encarnación F. 1985. Rearreglos cromosómicos y variabilidad cariotípica del género *Aotus* (Cebidae: Platyrrhini). *La Primatología en el Perú: Trabajos del Centro de Reproducción y Conservación de Primates. Tema* 4: 572-577.
- Dobigny G, Yang F, O'Brien PCM, Volobouev V, Kovács A, Pieczarka, JC, Ferguson-Smith MA, Robinson TJ. 2005. Low rate of genomic repatterning in Xenartha inferred from chromosome painting data. *Chromosome Research* 13(7): 651-663. <http://doi.org/10.1007/s10577-005-1002-9>.
- Duarte JMB, Giannoni ML. 1995. Cytogenetic analysis of the Marsh Deer, *Blastocerus dichotomus* (Mammalia: Cervidae). *Brazilian Journal of Genetics* 18: 245-248.
- Duarte JMB, Jorge W. 2003. Morphologic and cytogenetic description of the small red brocket (*Mazama bororo* Duarte, 1996) in Brazil. *Mammalia* 67(3): 403-410. <http://doi.org/10.1515/mamm.2003.67.3.403>.
- Dumas F, Bigoni F, Stone G, Sineo L, Stanyon R. 2005. Mapping genomic rearrangements in titi monkeys by chromosome flow sorting and multidirectional *in situ* hybridization. *Chromosome Research* 13(1): 85-96. <http://doi.org/10.1007/s10577-005-7063-y>.
- Dutrillaux B, Lombard M, Carroll JB, Martin RD. 1988. Chromosomal affinities of *Callimico goeldii* (Platyrrhini) and characterization of a Y-autosome translocation in the male. *Folia Primatologica* 50(3-4): 230-236. <http://doi.org/10.1159/000156349>.
- Egozcue J, Egozcue V. 1966. The chromosome complement of the howler monkey (*Alouatta caraya*, Humboldt 1812). *Cytogenetic and Genome Research* 5(1-2): 20-27. <http://doi.org/10.1159/000129881>.
- Egozcue J, Egozcue MV. 1967. The chromosome complement of *Cebus albifrons* (Erleben 1777). *Folia Primatologica* 5(4): 285-294. <http://doi.org/10.1159/000161953>.
- Egozcue J, Perkins EM, Hagemenas F. 1968. Chromosomal evolution in marmosets, tamarins, and pinchés. *Folia Primatologica* 9(2): 81-94. <http://doi.org/10.1159/000155170>.
- Egozcue J, Perkins EM, Hagemenas F. 1969. The chromosomes of *Saguinus fuscicollis illigeri* (Pucheran, 1845) and *Aotus trivirgatus* (Humboldt, 1811). *Folia Primatologica* 10(1-2): 154-159. <http://doi.org/10.1159/000155194>.
- Egozcue J, Perkins EM, Hagemenas F, Ford DM. 1969. The chromosomes of some Platyrrhini (*Callicebus*, *Ateles* and *Saimiri*). *Folia Primatologica* 11(1-2): 17-27. <http://doi.org/10.1159/000155256>.
- Finotelo LF, Amaral PJ, Pieczarka JC, Oliveira EH, Pissinati A, Neusser M, Müller S, Nagamachi, CY. 2010. Chromosome phylogeny of the subfamily Pitheciinae (Platyrrhini: Primates) by classic cytogenetics and chromosome painting. *BMC Evolutionary Biology* 10(1): 189. <http://doi.org/10.1186/1471-2148-10-189>.
- Fiorillo BF, Sarria-Perea JA, Abril VV, Duarte JMB. 2013. Cytogenetic description of the Amazonian brown brocket *Mazama nemorivaga* (Cetartiodactyla: Cervidae). *Comparative Cytogenetics* 7(1): 25. <http://doi.org/10.3897/CompCytogen.v7i1.4314>.
- Franco-de-Sá JFO, Rosas FCW, Feldberg E. 2007. Cytogenetic study of the giant otter *Pteronura brasiliensis* Zimmermann 1780 (Carnivora, Mustelidae, Lutrinae). *Genetics and Molecular Biology* 30(4): 1093-1096. <http://doi.org/10.1590/S1415-4757200700060001>.
- Freitas L, Seuánez H. 1982. Chromosome heteromorphisms in *Cebus apella*. *Journal of Human Evolution* 11(2): 173-180. [http://doi.org/10.1016/S0047-2484\(82\)80050-X](http://doi.org/10.1016/S0047-2484(82)80050-X).
- García M, Borrell A, Mudry M, Egozcue J, Ponsá M. 1995. Prometaphase karyotype and restriction-enzyme banding in squirrel monkeys, *Saimiri boliviensis boliviensis* (Primates: Platyrrhini). *Journal of Mammalogy* 76(2): 497-503. <http://doi.org/10.2307/1382358>.
- García M, Miró R, Ponsá M, Egozcue J. 1980. Banding patterns of the chromosomes of a specimen of *Lagothrix lagothricha cana*. *Genetica* 54(2): 181-184. <http://doi.org/10.1159/000155839>.
- Hatanaka T, Tambasco AJ, Galetti Junior PM. 1998. Heterochromatin heterogeneity and chromosome heteromorphism in *Cerdocyon thous* (Mammalia: Canidae). *Genetics and Molecular Biology* 21(2): 227-231.
- Henderson AS, Warburton DW, Megraw-Ripley SM, Atwood KC. 1977. The chromosomal location of rDNA in selected lower primates. *Cytogenetic and Genome Research* 19(5): 281-302. <http://doi.org/10.1159/000130821>.
- Houck ML, Kingswood SC, Kumamoto AT. 2000. Comparative cytogenetics of tapirs, genus *Tapirus* (Perissodactyla, Tapiridae). *Cytogenetic and Genome Research* 89(1-2): 110-115. <http://doi.org/10.1159/000015587>.
- Jacinto PJHR, Santiloni V, Rosa PS, da Mota LSL, Jorge W. 2009. The karyotype of *Cabassou uncinatus* (Dasypodidae, Xenartha). *Caryologia* 62(1): 24-29.
- Jordano P, Galetti M, Pizo MA, Silva WR. 2006. Ligando frugivoria e dispersão de sementes à biologia da conservação. *Biologia da conservação: essências*. Editorial Rima, São Paulo, Brasil: 411-436.
- Jorge W. 2000. Mitotic and meiotic chromosome studies in silky anteater *Cyclopes didactylus* (Myrmecophagidae: Xenartha). *Cytobios* 101(397): 95-100.
- Jorge W, Best RC, Wetzel RM. 1985. Chromosome Studies on the Silky Anteater *Cyclopes didactylus* L. (Myrmecophagidae: Xenartha: Edentata). *Caryologia* 38(3-4): 325-329. <http://doi.org/10.1080/0087114.1985.10797756>.
- Jorge W, Meritt JD, Benirschke K. 1978. Chromosome studies in Edentata. *Cytobios* 18(71-72): 157-172.
- Koiffman CP, Saldanha PH. 1978. The karyotype of *Brachyteles arachnoides* (E. Geoffroy, 1806) (Primates: Platyrrhini). *Genetica* 48(2): 129-130. <http://doi.org/10.1007/BF00127508>.
- Koiffmann CP, Saldanha PH. 1981. The karyotype of *Cacajao melanocephalus* (Platyrrhini, Primates). *Folia Primatologica* 36(1-2): 150-154. <http://doi.org/10.1159/000156015>.
- Langguth A, Sousa MAN. 2008. The karyotype of *Sylvilagus brasiliensis minensis* Thomas (Lagomorpha: Leporidae). *Revista Nordestina de Biologia* 17(1): 59-62.



- Ledesma MA, Ledesma CO, Schiaffino K, Rinas MA, Gunski RJ. 2004. Análisis citogenético de *Panthera onca* (Felidae: Pantherinae) de la provincia de Misiones, Argentina. *Mastozoología Neotropical* 1(11): 85-90. <http://www.redalyc.org/pdf/457/45711109.pdf>.
- Lima JFDS, Guedes FB, Silva RW, Hass I, Cavalli JJ, Silva JD, de Freitas TR, Sbalqueiro JJ. 2004. Unexpected chromosomal alterations in *Tayassu tajacu* (Cetartiodactyla: Tayassuidae) in captivity. *Brazilian Journal of Veterinary Research and Animal Science* 41(1): 10-13. <http://doi.org/10.1590/S1413-95962004000100002>.
- Lima MMC, Seuánez HN. 1989. Cytogenetic characterization of *Alouatta belzebul* with atypical pelage coloration. *Folia Primatologica* 52(1-2): 97-101. <http://doi.org/10.1159/000156387>.
- Lima MMC, Seuánez HN. 1991. Chromosome studies in the red howler monkey, *Alouatta seniculus stramineus* (Platyrrhini, Primates): description of an X1X2Y1Y2/X1X1X2X2 sex-chromosome system and karyological comparisons with other subspecies. *Cytogenetic and Genome Research* 57(2-3): 151-156. <http://doi.org/10.1159/000133135>.
- Lizarralde, MS, Bolzán, AD, Poljak, S, Pigozzi, MI, Bustos, J, Merani, MS. 2005. Chromosomal localization of the telomeric (TTAGGG) n sequence in four species of armadillo (Dasypodidae) from Argentina: an approach to explaining karyotype evolution in the Xenarthra. *Chromosome Research* 13(8), 777-784. <http://doi.org/10.1007/s10577-005-1011-8>.
- Ma NS, Jones TC, Miller AC, Morgan LM, Adams EA. 1976. Chromosome polymorphism and banding patterns in the owl monkey (*Aotus*). *Laboratory Animal Science* 26(6 Pt 2): 1022-1036.
- Ma NS, Renquist DM, Hall R, Sehgal PK, Simeone T, Jones TC. 1980. XX/"XO" sex determination system in a population of Peruvian owl monkey, *Aotus*. *Journal of Heredity* 71(5): 336-342. <http://doi.org/10.1093/oxfordjournals.jhered.a109382>.
- Marczynska B, Peterson DA, Ogdén JD, Wolfe LG. 1983. Karyotype of *Saguinus labiatus labiatus* (red-bellied marmosets). *Folia Primatologica* 40(3): 217-226. <http://doi.org/10.1159/000156102>.
- Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J, Shago M, Moessner R, Pinto D, Ren Y, Thiruvahindrapuram B, Fiebig A, Schreiber S, Friedman J, Ketelaars CEJ, Vos YJ, Ficioglu C, Kirkpatrick S, Nicolson R, Sloman L, Summers A, Gibbons CA, Teebi A, Chitayat D, Weksberg R, Thompson A, Vardy C, Crosbie V, Luscombe S, Baatjes R, Zwaigenbaum L, Roberts W, Fernandez B, Szatmari P, Scherer SW. 2008. Structural variation of chromosomes in autism spectrum disorder. *American Journal of Human Genetics* 82(2): 477-488. <http://doi.org/10.1016/j.ajhg.2007.12.009>.
- Medeiros MA, Barros RMS, Pieczarka JC, Nagamachi CY, Ponsa M, Garcia M, Garcia F, Egozcue J. 1997. Radiation and speciation of spider monkeys, genus *Ateles*, from the cytogenetic viewpoint. *American Journal of Primatology* 42(3): 167-178. [http://doi.org/10.1002/\(SICI\)1098-2345\(1997\)42:3<167::AID-AJP1>3.0.CO;2-V](http://doi.org/10.1002/(SICI)1098-2345(1997)42:3<167::AID-AJP1>3.0.CO;2-V).
- Menezes AN, Bonvicino CR, Seuánez HN. 2010. Identification, classification and evolution of owl monkeys (*Aotus*, Illiger 1811). *BMC Evolutionary Biology* 10(1): 248. <http://doi.org/10.1186/1471-2148-10-248>.
- Mudry MD, Slavutsky I, de Vinuesa ML. 1990. Chromosome comparison among five species of Platyrrhini (*Alouatta caraya*, *Aotus azarae*, *Callithrix jacchus*, *Cebus apella*, and *Saimiri sciureus*). *Primates* 31(3): 415-420. <http://doi.org/10.1007/BF02381112>.
- Nagamachi CY, Pieczarka JC. 1988. Chromosome studies of *Saguinus midas niger* (Callitrichidae, Primates) from Tucuruí, Para, Brazil: comparison with the karyotype of *Callithrix jacchus*. *American Journal of Primatology* 14(3): 277-284. <http://doi.org/10.1002/ajp.1350140308>.
- Nagamachi CY, Pieczarka JC, Barros RMS, Schwarz M, Muniz JA, Mattevi MS. 1996. Chromosomal relationships and phylogenetic and clustering analyses on genus *Callithrix*, group *argentata* (Callitrichidae, Primates). *Cytogenetic and Genome Research* 72(4): 331-338. <http://doi.org/10.1159/000134216>.
- Nagamachi CY, Pieczarka JC, Schwarz M, Barros RM, Mattevi MS. 1997. Comparative chromosomal study of five taxa of genus *Callithrix*, group *jacchus* (Platyrrhini, Primates). *American Journal of Primatology* 41(1): 53-60. [http://doi.org/10.1002/\(SICI\)1098-2345\(1997\)41:1<53::AID-AJP5>3.0.CO;2-Z](http://doi.org/10.1002/(SICI)1098-2345(1997)41:1<53::AID-AJP5>3.0.CO;2-Z).
- Nagamachi CY, Pieczarka JC, Schwarz M, Paiva C, Barros R, Mattevi MS. 1994. Karyotype of *Callithrix mauesi* (Callitrichidae, Primates) and its relations with those of *C. emiliae* and *C. jacchus*. *American Journal of Primatology* 33(4): 309-315. <http://doi.org/10.1002/ajp.1350330405>.
- Nagamachi CY, Rodrigues LR, Galetti Jr PM, Mantovani M, Pissinati A, Rissino JD, Barros RMS, Pieczarka, JC. 2003. Cytogenetic studies in *Callicebus personatus nigrifrons* (Platyrrhini, Primates). *Caryologia* 56(1): 47-52. <http://doi.org/10.1080/00087114.2003.10589306>.
- Neitzel H. 1979. Chromosome evolution in der Familie der Hirsche (Cervidae). *Bongo* 3: 27-38.
- Oliveira C, Foresti F, Hilsdorf AWS. 2009. Genetics of neotropical fish: from chromosomes to populations. *Fish Physiology and Biochemistry* 35(1): 81-100. <http://doi.org/10.1007/s10695-008-9250-1>.
- Oliveira EC, Suemitsu E, Silva A, Sbalqueiro JJ. 2000. Geographical variation of chromosomal number in *Alouatta fusca damitans* (Primates, Ateleidae). *Caryologia* 53(2): 163-168. <http://doi.org/10.1080/00087114.2000.10589192>.
- Oliveira EH, Lima M, Sbalqueiro JJ, Pissinati A. 1998. The karyotype of *Alouatta fusca clamitans* from Rio de Janeiro, Brazil: Evidence for a y-autosome translocation. *Genetics and Molecular Biology* 21(3). <http://doi.org/10.1590/S1415-47571998000300012>.
- Paglia AP, da Fonseca GA, Rylands AB, Herrmann G, Aguiar LM, Chiarello AG, Mendes SL. 2012. Lista Anotada dos Mamíferos do Brasil 2ª Edição/Annotated Checklist of Brazilian Mammals 2nd Edition. *Occasional Papers in Conservation Biology* 6: 1-76.
- Paresque R, Rodrigues JS, Riguetti KB. 2018. Karyotypes of Brazilian non-volant small mammals (Didelphidae and Rodentia): An online tool for accessing the chromosomal diversity. *Genetics and Molecular Biology* 41(3): 605-610.
- Pargament MM, Colillas OJ, de Salum SB. 1984. The *Aotus* from northern Argentina. *Primates* 25(4): 530-537.
- Percequillo AR, Hingst-Zaher E, Bonvicino CR. 2008. Systematic review of genus *Cerradomys* Wexler, Percequillo and Voss, 2006 (Rodentia: Cricetidae: Sigmodontinae: Oryzomyini), with description of two new species from eastern Brazil. *American Museum Novitates* 1-46. <http://doi.org/10.1206/495.1>.
- Pereira Júnior HRJ, Jorge W, Costa MELTD. 2004. Chromosome study of *Anteaters* (Myrmecophagidae, Xenarthra): a preliminary report. *Genetics and Molecular Biology* 27(3): 391-394. <http://doi.org/10.1590/S1415-47572004000300014>.
- Pieczarka JC. 1989. The karyotype of *Ateles paniscus paniscus* (Cebidae, Primates): 2n = 32. *Revista Brasileira de Genética* 12: 543-551.
- Pieczarka JC, Barros RMS, Nagamachi CY, Rodrigues R, Espinel A. 1992. *Aotus vociferans* × *Aotus nancymai*: Sympatry without chromosomal hybridation. *Primates* 33(2): 239-245. <http://doi.org/10.1007/BF02382753>.
- Pieczarka JC, Nagamachi CY. 1988. Cytogenetic studies of *Aotus* from Eastern Amazonia. Y/Autosome rearrangement. *American Journal of Primatology* 14(3): 255-263. <http://doi.org/10.1002/ajp.1350140306>.
- Pieńkowska-Schelling A, Schelling C, Zawada M, Yang F, Bugno, M, Ferguson-Smith M. 2008. Cytogenetic studies and karyotype nomenclature of three wild canid species: maned wolf (*Chrysocyon brachyurus*), bat-eared fox (*Otocyon megalotis*) and fennec fox (*Fennecus zerda*). *Cytogenetic and Genome Research* 121(1): 25-34. <http://doi.org/10.1159/000124378>.
- Redi CA, Zacharias H, Merani S, Oliveira-Miranda M, Aguilera M, Zuccotti M, Garagna S, Capanna E. 2005. Genome sizes in Afrotheria, Xenarthra, Euarchontoglires, and Laurasiatheria. *Journal of Heredity* 96(5): 485-493. <http://doi.org/10.1093/jhered/esi080>.
- Robinson TJ, Yang F. 2012. Molecular cytogenetics: karyotype evolution, phylogenomics and future prospects. *Heredity* 108: 1-3. <http://doi.org/10.1038/hdy.2011.117>.
- Rodrigues LRR, Barros RMS, Pissinati A, Pieczarka JC, Nagamachi CY. 2001. Cytogenetic study of *Callicebus hoffmannsi* (Cebidae, Primates) and comparison with *C. m. moloch*. *Cytobios* 105: 137-145. <http://doi.org/10.1080/00087114.2003.10589306>.
- Rodrigues LRR, Barros R, Pissinati A, Pieczarka JC, Nagamachi CY. 2004. A new karyotype of an endangered primate species (*Callicebus personatus*) from the Brazilian Atlantic forests. *Hereditas* 140(2): 87-91. <http://doi.org/10.1111/j.1601-5223.2004.01793.x>.
- Rodrigues LRR, Sousa MC, Pieczarka JC, Nagamachi CY. 2006. Karyotypic study of *Callicebus coimbrai*: a rare and threatened primate species from Brazil. *Caryologia* 59(3): 248-252.
- Romagno D. 2001. Primate tables chromosome. *Caryologia* 54(4), 285-297.



- Romanenko, SA, Volobouev, VT, Perelman, PL, Lebedev, VS, Serdukova, NA, Trifonov, VA, Biltueva LS, Nie, W, O'Brien PCM, Bulatova NS, Ferguson-Smith, MA. 2007. Karyotype evolution and phylogenetic relationships of hamsters (Cricetidae, Muroidea, Rodentia) inferred from chromosomal painting and banding comparison. *Chromosome Research* 15(3): 283-298. <http://doi.org/10.1007/s10577-007-1124-3>.
- Saez FA, Drets ME, Brum N. 1964. The chromosomes of the mulita (*Dasyopus hybridus*): a mammalian edentate of South America. *Mammalian Cytogenetics and Related Problems in Radiobiology* 163-170.
- Sagrillo MR, Cardoso SH, Silva LR, Graça CH, Ferreira E, Hamerschlag N, Guerra JCC, Bacal NS, Andrade JAD, Borovik CL. 2005. Leucemia promielocítica aguda: caracterização de alterações cromossômicas por citogenética tradicional e molecular (FISH). *Revista Brasileira de Hematologia e Hemoterapia* 27(2): 94-101. <http://doi.org/10.1590/S1516-84842005000200008>.
- Sazima I, Bokermann WCA. 1982. Anfíbios da Serra do Cipó, Minas Gerais, Brasil. 5: *Hylodes otavioi* sp. n. (Anura, Leptodactylidae). *Revista Brasileira de Biologia* 42(4): 767-771.
- Sbalqueiro IJ, Persson VG, Lorini ML. 1992. O cariótipo de *Leontopithecus caissara* (Callitrichidae, Primates). Pp. 172, In: Anais do XIX Congresso Brasileiro de Zoologia. SBZ, Belém.
- Seuáñez HN, Alves G, Lima MMC, Barros RDS, Barros CML, Muniz JAPC. 1992. Chromosome studies in *Chiropotes satanas utahicki* Hershkovitz, 1985 (Cebidae, Platyrrhini): A comparison with *Chiropotes satanas chiropotes*. *American Journal of Primatology* 28(3): 213-222. <http://doi.org/10.1002/ajp.1350280306>.
- Seuáñez HN, Forman L, Alves G. 1988. Comparative chromosome morphology in three callitrichid genera: *Cebuella*, *Callithrix*, and *Leontopithecus*. *Journal of Heredity* 79(6): 418-424.
- Sherlock JK, Griffin DK, Delhanty JDA, Parrington JM. 1996. Homologies between human and marmoset (*Callithrix jacchus*) chromosomes revealed by comparative chromosome painting. *Genomics* 33(2): 214-219. <http://doi.org/10.1006/geno.1996.0186>.
- Stanyon R, Bonvicino CR, Svartman M. 2003. Chromosome painting in *Callicebus lugens*, the species with the lowest diploid number (2n = 16) known in primates. *Chromosoma* 112(4): 201-206. <http://doi.org/10.1007/s00412-003-0261-5>.
- Stanyon R, Consigliere S, Bigoni F, Ferguson-Smith M, O'Brien P, Wienberg J. 2001. Reciprocal chromosome painting between a New World primate, the woolly monkey, and humans. *Chromosome Research* 9(2): 97-106. <http://doi.org/10.1023/A:1009274802086>.
- Stanyon R, Consigliere S, Müller S, Morescalchi A, Neusser M, Wienberg J. 2000. Fluorescence *in situ* hybridization (FISH) maps chromosomal homologies between the dusky titi and squirrel monkey. *American Journal of Primatology* 50(2): 95-107. [http://doi.org/10.1002/\(SICI\)1098-2345\(200002\)50:2<95::AID-AJP1>3.0.CO;2-8](http://doi.org/10.1002/(SICI)1098-2345(200002)50:2<95::AID-AJP1>3.0.CO;2-8).
- Viana MC, Bonvicino CR, Ferreira JG, Jerusalinsky L, Langguth A, Seuáñez H. 2015. Understanding the relationship between *Alouatta ululata* and *Alouatta belzebul* (Primates: Atelidae) based on cytogenetics and molecular phylogeny. *Oecologia Australis* 19: 173-182.
- Voss, RS, Jansa, SA. 2009. Phylogenetic relationships and classification of didelphid marsupials, an extant radiation of New World metatherian mammals. *Bulletin of the American Museum of Natural History* 1-177. <http://doi.org/10.1206/322.1>.
- Wurster DH, Benirschke K. 1967. Chromosome studies in some deer, the springbok, and the pronghorn, with notes on placentation in deer. *Cytologia* 32(2): 273-285. <http://doi.org/10.1508/cytologia.32.273>.
- Wurster DH, Benirschke K. 1968. Comparative cytogenetic studies in the order Carnivora. *Chromosoma* 24(3): 336-382. <http://doi.org/10.1007/BF00336201>.
- Wurster DH, Benirschke K. 1969. Chromosomes of some primates. *Mammalian Chromosomes Newsletter* 10(3).
- Yonenaga-Yassuda Y, Chu TR. 1985. Chromosome banding patterns of *Saimiri vanzolinii* Ayres, 1985 (Primates, Cebidae). *Papéis Avulsos de Zoologia* 36: 165-168.
- Yunis EJ, De Caballero OMT, Ramírez C, Ramírez E. 1976. Chromosomal variations in the primate *Alouatta seniculus*. *Folia Primatologica* 25(2-3): 215-224. <http://doi.org/10.1159/000155714>.

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